



US005194300A

**United States Patent** [19]  
**Cheung**[11] **Patent Number:** **5,194,300**  
[45] **Date of Patent:** **Mar. 16, 1993**[54] **METHODS OF MAKING FLUORESCENT MICROSPHERES**[76] **Inventor:** **Sau W. Cheung, 8528 Douglas Ct., St. Louis, Mo. 63144**[21] **Appl. No.:** **433,677**[22] **Filed:** **Nov. 8, 1989****Related U.S. Application Data**[62] **Division of Ser. No. 73,770, Jul. 15, 1987, abandoned.**[51] **Int. Cl.<sup>3</sup>** ..... **G01N 33/547; C08F 8/10**[52] **U.S. Cl.** ..... **427/213.31; 525/54.1; 252/301.35; 436/533; 436/532; 436/546; 436/800; 428/407; 530/402; 530/816**[58] **Field of Search** ..... **427/213.3, 213.31, 213.34; 428/402.24; 436/533, 534; 525/54.1, 54.11; 252/301.36, 301.35**[56] **References Cited****U.S. PATENT DOCUMENTS**

3,853,987	12/1974	Dreyer	.....	424/1.1
4,035,316	7/1977	Yen et al.	.....	436/546
4,105,598	8/1978	Yen et al.	.....	428/402 X
4,108,972	8/1978	Dreyer	.....	436/534
4,138,383	2/1979	Rembaum et al.	.....	525/54.1 X
4,140,662	2/1979	Reckel et al.	.....	436/533
4,143,203	3/1979	Rigopulos et al.	.....	436/533 X
4,224,198	9/1980	Rembaum et al.	.....	525/54.1
4,326,008	4/1982	Rembaum	.....	428/403
4,438,239	3/1984	Rembaum et al.	.....	525/54.1
4,489,055	12/1984	Couvreur et al.	.....	427/213.31 X
4,510,244	4/1985	Parks et al.	.....	436/533
4,522,812	11/1985	Margel et al.	.....	427/213.34
4,622,362	11/1986	Rembaum	.....	525/54.1
4,699,826	10/1987	Schwartz et al.	.....	525/54.1 X
4,735,907	4/1988	Schaeffer et al.	.....	436/533 X
4,783,336	11/1988	Margel et al.	.....	427/213.34 X
4,837,168	6/1989	de Jaeger et al.	.....	436/533

**FOREIGN PATENT DOCUMENTS**

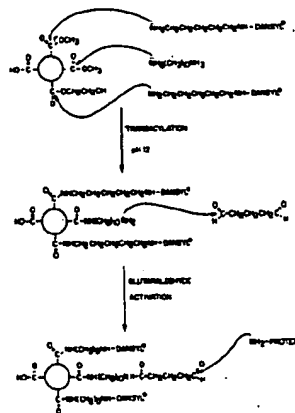
1124643 4/1982 Canada .

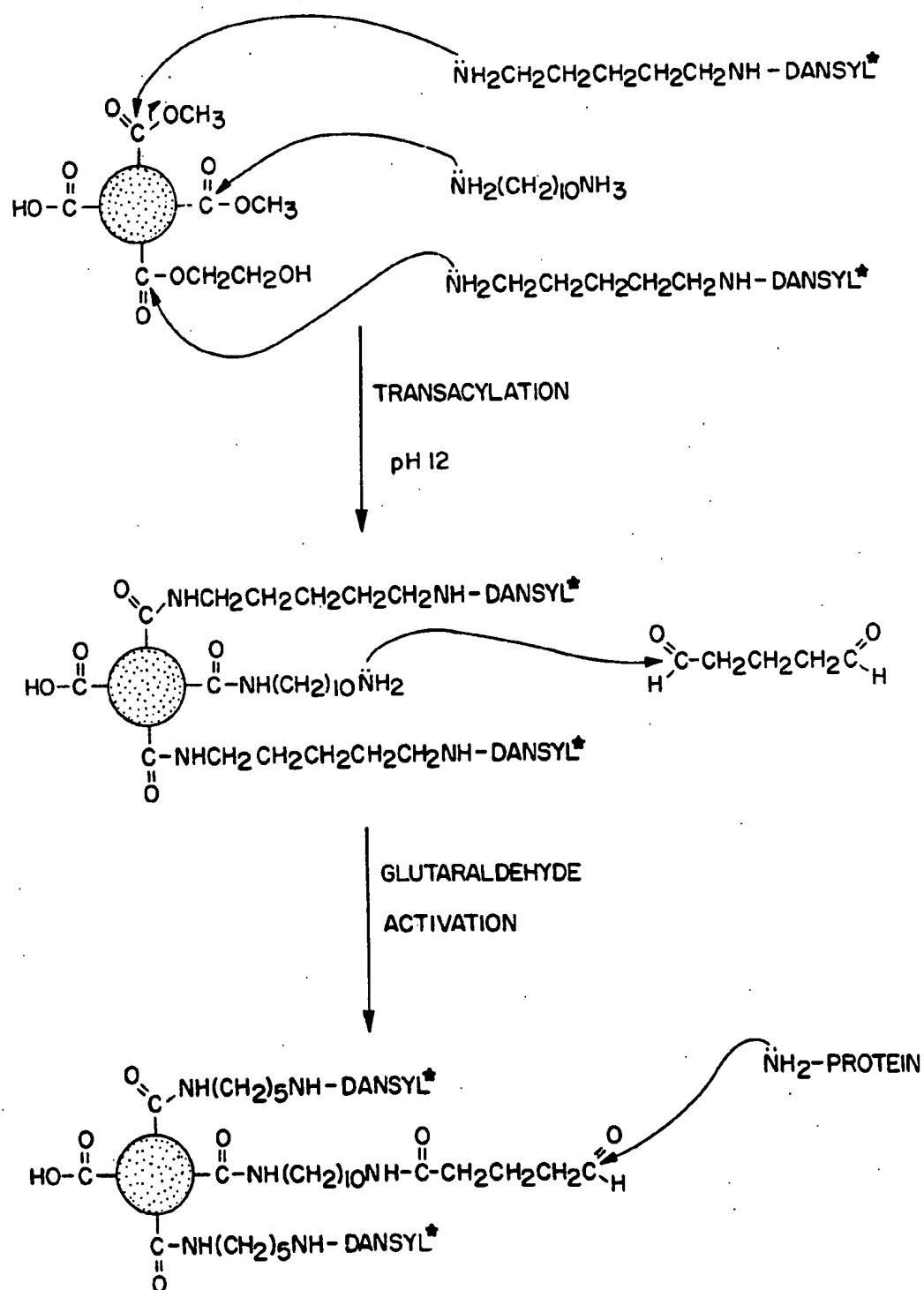
**OTHER PUBLICATIONS**Cheung et al., *Cell Biology International Reports*, vol. 1, No. 3, 1977, pp. 255-262.Klausner et al., *Bio/Technology*, Aug. 1983, pp. 471-478.Nairn (ed), *Fluorescent Protein Tracing*, Churchill Livingstone Ltd., 1976, Chpt. 4, pp. 68-107.Muirhead et al., *Bio/Technology*, vol. 3, Apr. 1985, pp. 337-356.R. S. Molday, W. J. Dreyer, A. Rembaum, and Yen, *Journal of Cell Biology*, vol. 64, 1975, pp. 75-88.Kaplan et al., *Biochimica et Biophysica Acta*, vol. 728, 1983, pp. 112-120.Parks et al., *Genetics*, vol. 76, No. 4, Apr. 1979, pp. 1962-1966.Fuccillo, *Bio Techniques*, vol. 3, No. 6, 1985, pp. 494-501.Fornusek et al., *CRC Critical Reviews in Therapeutic Drug Carrier Systems*, vol. 2, Issue 2, 1986, pp. 137-174.Rembaum and Dreyer, *Science*, vol. 208, 25 Apr. 1980, pp. 364-368.Feinberg et al., *Analytical Biochemistry* 132, 1983, pp. 6-13.Harper et al., *Chromosoma*, 83, 1981, pp. 431-439.March, *Advanced Organic Chemistry: Reactions, Mechanisms and Structure*, McGraw-Hill, N.Y., 2nd edition, 1977, pp. 386-387.

Cheung et al., (abstract) Annual Meeting, American Society of Human Genetics, San Diego, Oct. 19-22, 1977.

Cheung et al., *Advances in Gene Technology: The Molecular Biology of Development*, ICSU Short Reports, vol. 7, 1987, p. 123.*Primary Examiner*—Robert L. Stoll*Assistant Examiner*—John M. Covert*Attorney, Agent, or Firm*—Polster, Lieder, Woodruff & Lucchesi[57] **ABSTRACT**

Highly fluorescent latex microspheres have a diameter of less than five hundred angstroms and have more than five thousand fluorescent markers per sphere. The microspheres are prepared by reacting an acrylic latex bead with a diamine and a fluorescent amine at elevated pH. A protein such as avidin or an immunoglobulin may then be conjugated to the diamine. A single fluorescent microsphere is visible using standard fluorescent microscopy. Therefore the microspheres may be utilized not only to visualize cell surface antigens but also DNA encoding for single genes, by means of a biotinylated DNA probe.

**20 Claims, 1 Drawing Sheet**



## METHODS OF MAKING FLUORESCENT MICROSPHERES

### CROSS-REFERENCE TO RELATED APPLICATION

This is a division of copending application Ser. No. 07/073,770, filed Jul. 15, 1987, now abandoned.

### BACKGROUND OF THE INVENTION

This invention relates to microspheres and particularly to fluorescent microspheres and methods of making and using them. Fluorescent microspheres have long been known, but their use has been limited by the complexity of making them, their limited visibility, the limited stability of some forms of them, their tendency to clump together, the lack of sensitivity of procedures using them, and their tendency to produce a large background fluorescence which makes visualization difficult and causes diagnostic error.

Biochemical probes have found widespread use in biochemistry and medicine. A popular review of these technologies is found in Klausner and Wilson, "Gene Detection Technology Opens Doors for Many Industries," *Bio/Technology*, August 1983, pp. 471-478. The most popular of these probes include a radioactive tracer, or an enzyme which catalyzes color-producing reactions, or a fluorescent species (fluorochrome) which fluoresces brightly under light of the appropriate wavelength. As set out in that article, some of these techniques presently permit detection of as few as 10<sup>4</sup> copies of a single gene, although the techniques require several processing steps and several hours to carry out.

Radioactive probes create substantial handling and waste disposal problems, and color-forming enzymes are insensitive and hence require a large sample of the substrate to visualize.

Fluorochrome labels have now found wide use in microbiology. They are particularly useful in localizing cell surface antigens and in detecting specific nucleic acid sequences, although their usefulness is by no means limited to these applications. Two well-known methods of detecting fluorochrome-labeled species are fluorescence microscopy, described for example in *Fluorescent Protein Tracing*, ed. by R. C. Nairn, especially Chapter 4 (Churchill Livingstone Ltd. 1976), and flow cytometry, described for example in Muirhead et al, *Bio/Technology*, 3, pp. 337-356 (1985).

Direct staining with fluorescently-labeled antibodies presents several difficulties, such as the complication of the procedure and its tendency to inactivate the antibody during attachment of the fluorescent marker. These difficulties are partially overcome by a sandwich technique in which a fluorescent marker is attached to a secondary antibody that in turn binds to a specific primary antibody. The lack of efficiency and specificity of the secondary binding, however, introduces new problems with this type of indirect method.

A "fluorescent vehicle" for attachment as a visible marker to an IgG fraction of secondary antibody was developed by Molday, Dreyer, Rembaum, and Yen, *Journal of Cell Biology*, 64, pp. 75-88 (1975). By emulsion copolymerization they synthesized latex microspheres having a diameter from 300 angstroms to 3400 angstroms. These microspheres are designed to be attached to a substrate, namely cell surface antigens to which primary antibodies have been attached, via secondary antibodies bound to the spheres. The spheres

may be visualized via electron microscopy or by the addition to the spheres of radioactive or fluorochrome probes. The microspheres contain about 57% methyl ester terminations, about 30% hydroxyethyl ester groups, and about 10% carboxylic acid groups. Fluorescent species were attached to some of these microspheres, having a diameter of 600 angstroms, through the hydroxyethyl ester groups via a cyanogen bromide reaction, and the secondary antibody was conjugated to the carboxylic acid groups. The spheres had limited fluorescence and tended to clump into large aggregates. The spheres were used in an indirect method of localizing cell surface antigens.

Kaplan et al, *Biochimica et Biophysica Acta*, 728, pp. 112-120 (1983) modified this technique and synthesized 500 angstrom diameter microspheres containing a fluorescent cross-linking agent, fluorescein. This technique allowed them to incorporate 700 fluorescein molecules per microsphere and reduced background fluorescence. The fluorescent microspheres were then conjugated to acetylated avidin which binds to biotinylated monoclonal antibody using both the sandwich method and direct detection of surface antigens. This involves several steps in addition to the synthesis of fluorescent microspheres, including acetylation of avidin, purification of monoclonal antibody and biotinylation of monoclonal antibody. However, this technique demonstrated the fact that fewer than ten thousand receptors per cell can be detected by fluorescent flow cytometry. These workers suggested the use of larger spheres, which enable them to attach more fluorochrome and thereby enhance the fluorescent signal, to detect cells containing even fewer receptors. Such an approach would also decrease sensitivity because of steric hindrance.

Parks et al, *Proc. Natl. Acad. Sci. (USA)*, 76, pp. 1962-66 (1979), using 0.783 micrometer (7830 angstrom) fluorescent microspheres, showed that as few as some tens of microsphere-antigen conjugates per cell could be used to select hybridomas from mixtures. These authors report that individual beads were visible under laser excitation.

Fuccillo, *BioTechniques*, 3, pp. 494-501 (1985) describes other uses of the avidin-biotin complex.

Fornusek and Vetvicka, "Polymeric Microspheres as Diagnostic Tools for Cell Surface Marker Tracing," in *CRC Critical Reviews in Therapeutic Drug Carrier Systems*, 2, pp. 137-174 (1986), includes an extensive review of the use of microspheres, including fluorescent microspheres, for cell surface marking. These authors point out that, "Despite all the progress it has brought to cell biology, the fluorescence detection of cell surface markers is relatively tedious and rather insensitive with respect to the recent applications of it." These authors utilize fluorescent microspheres which are on the order of 20,000 angstroms, and cite others who utilize microspheres (Covaspheres™) of about 7,000 to 9,000 angstroms in diameter to make the microspheres individually observable.

### SUMMARY OF THE INVENTION

One of the objects of this invention is to provide a method of forming microspheres which have a far greater degree of fluorescence than previously known fluorescent microspheres.

Another object of this invention is to provide such microspheres which may be individually observable by

either manual or automated fluorescence microscopy, but which are quite small.

Another object of this invention is to provide such microspheres which do not have a tendency to clump and which will remain in single suspension and will react individually with substrates.

Another object of this invention is to provide such microspheres which provide a sharp contrast between background and the substrate to which they are bound.

Another object is to provide methods of using such microspheres to increase the sensitivity and resolution of known fluorescence microscopy and flow cytometry procedures and to detect features of substrates which heretofore were not detectable, such as precise topological resolution of antigenic sites and rapid, high-resolution mapping of single gene sequences.

Other objects of this invention will be apparent to those skilled in the art in light of the following description and accompanying drawings.

In accordance with one aspect of this invention, generally stated, a method of forming a fluorescent bead is provided which comprises simultaneously reacting an acrylic latex bead with a fluorescent amine and with a diamine to form a conjugate of the fluorescent amine and the diamine with the bead, and thereafter a step of separating the bead from unconjugated fluorescent amine and unconjugated diamine. The method is preferably carried out at a high pH, most preferably a pH of 11 to 13.5. The diamine is preferably a straight chain aliphatic having a length of from eight to twenty carbon atoms. A protein, such as avidin or an immunoglobulin, is conjugated to the free end of the diamine. The fluorescent amine is preferably a straight chain aliphatic having a length of from three to seven carbon atoms and a fluorescent dansyl moiety, but any fluorochrome with a primary amino group may be attached to the spheres. Preferably, the fluorescent amine and the diamine are present in the reaction in a ratio of from three to five parts of fluorescent amine to one part diamine.

The latex bead of the invention is preferably made by a modification of the method of Molday et al, supra. The untreated bead preferably has both alkoxy groups and alkoxy alcohol groups, both of which react with the fluorescent amine and the diamine reactants. In this way, a far greater percent of the bead is covered with fluorescent moieties than in the Molday et al procedure, and the relative amounts of fluorescent amine markers and diamine attachment points is controlled more precisely.

The microsphere of the present invention is a latex bead having a diameter of less than 500 angstroms, having conjugated to its surface at least 5,000 fluorescent molecules. A plurality of these beads may be used as a marker system of great sensitivity and precision. The beads are preferably acrylic beads having a diameter of from 200 to 400 angstroms and preferably have in excess of 10,000 fluorescent molecules conjugated per bead. Preferably, a protein such as avidin or an immunoglobulin is conjugated to the bead through a spacer containing a chain of from eight to twenty carbon atoms. The fluorescent molecule is preferably conjugated to the bead through an amine linkage and a straight chain aliphatic having a length of from three to seven carbon atoms.

The intensely fluorescent, approximately 300 angstrom diameter microsphere of the invention may be utilized in either manual or automated fluorescence microscopy to determine the precise location of the

binding site of the microsphere on a substrate. For example, at a magnification of 2500X, under ultraviolet light, individual microspheres of the preferred embodiment are easily visible, emitting light at 430 mμ and 525 mμ. At this magnification, the location of a microsphere adhered to a human chromosome may easily be determined. At a magnification of 26,800X, electron photomicrographs of the microspheres show their location on DNA-biotin-avidin-microsphere complexes.

Other aspects of the invention will best be understood in light of the following description of the preferred embodiments.

### BRIEF DESCRIPTION OF THE DRAWINGS

In the drawings, the FIGURE is a diagrammatic representation of a method of forming a fluorescent bead in accordance with the present invention.

### DESCRIPTION OF THE PREFERRED EMBODIMENT

The following examples are illustrative of the methods and microspheres of the present invention.

#### EXAMPLE 1

##### Production of Microspheres

Latex microspheres having a mean diameter of 300 angstroms were prepared by a modification of the emulsion polymerization method of Molday, et al, *Journal of Cell Biology*, 64, pp. 75-88 (1975). Briefly, four acrylic monomers obtained from Fluka Chemical Company were purified in the following manner:

2-Hydroxyethyl methacrylate (HEMA)—distilled in the presence of a 2% hydroquinone at 76°C, 1.1 mm Hg pressure.

Methyl methacrylate (MMA)—distilled in the presence of a 0.5% hydroquinone at 57°C, 150 mm Hg pressure.

Methylacrylic acid (MAA)—distilled with 0.05% hydroquinone at 63°C, 10 mm Hg pressure.

Ethylene glycol dimethacrylate (EGD)—distilled with 2% hydroquinone at 86°C and 1.1 mm Hg pressure.

A suspension of microspheres was made by adding 0.9 parts by weight of HEMA, 1.71 parts of MMA, 0.3 parts MAA, 0.09 parts EGD, 0.120 parts of sodium dodecyl sulfate (SDS—an emulsifying agent), and 0.013 parts of ammonium persulfate (AP— a cross-linking agent) to 100 parts of distilled water. The emulsion was rotated in a specially constructed tumbling container at 98° C., under argon gas for 1 hour. Emulsifier and other ionic impurities were removed from the resultant suspension by dialysis in an Amicon ultrafiltration chamber until the conductivity of the filtrate was less than 5 μmho.

The synthetic microspheres were examined by transmission electron microscopy. They appeared as uniformly round bodies ranging from 200 to 400 angstroms in diameter. The presence of hydroxyethyl ester, carboxy, and methyl ester functional groups enables molecules containing primary amino groups to be covalently bonded to the microspheres by an assortment of chemical reactions. The resulting 300 angstrom diameter spheres have surfaces containing about 10% carboxylic acid terminations, about 30% hydroxyethyl ester terminations, and about 57% methyl ester terminations.

## EXAMPLE 2

## Production of Fluorescent Microspheres

The acrylic latex spheres of Example 1 were provided both with fluorescent markers and with spacers in a single reaction mixture by reacting not only the methyl ester groups but also the hydroxyethyl ester groups with a mixture of 80% dansylcadaverine (1-dansyl, 1,5-diamino pentane) and 20% 1,12-diaminododecane. A mixture of 30 mg of microspheres, 8 mg of fluorescent amine, and 2 mg of diamine were suspended in 50 milliliters of water. The pH was adjusted to 12 with 1.0N NaOH, and the mixture was allowed to stir overnight at room temperature. Under these reaction conditions, the methyl ester and hydroxyethyl ester groups can be replaced by forming covalent bonds with the amine moieties through a transacylation reaction as described in March, *Advanced Organic Chemistry: Reactions, Mechanisms, and Structure*, 2nd edition 1977, pp. 386-87. As set out in this reference, the conversion of esters to amides is a useful reaction, and unsubstituted, N-substituted, and N,N-disubstituted amides can be prepared in this way from the appropriate amine. The ester group may be alkyl or aryl.

The resulting microspheres were dialyzed against deionized water, pH 9.0 for 5 days, changed two to three times daily to remove the unconjugated dansyl cadaverine and diamine.

The resulting microspheres remain in single suspension, without clumping.

The extent of conjugation was estimated by incorporation of radioactive ethanolamine under the foregoing conditions. The number of covalently bound dansylcadaverine markers was estimated to be 0.005 micromoles per milliliter of solution.

The average size of the microspheres was determined by electron microscopy to be 310 angstroms in diameter, and the number of dansylcadaverine markers covalently attached per microsphere was determined by calculation to be approximately 14,400. This corresponds to about 5,000,000 molecules of dansylcadaverine per square micrometer on the surface of the average fluorescent microsphere. This is far above the minimum of 1,000 molecules per square micrometer generally regarded as necessary for visualizing microspheres by fluorescent microscopy. The density of fluorescent moieties bound to the microsphere is many times that of previously known microspheres.

The microspheres were found to be individually easily observable by fluorescent microscopy at a magnification of 630 $\times$  to 1,000 $\times$ .

The resulting fluorescent microspheres were found to be remarkably stable for periods of more than one year.

## EXAMPLE 3

## Attachment of Antibody to Fluorescent Microspheres

Fluorescent microspheres produced in accordance with Example 2 were covalently conjugated to the well-established anti-H-2K monoclonal antibody following a two step glutaraldehyde reaction in accordance with the method described by Rembaum and Dreyer, *Science*, 208, pp. 364-68 (1980). The activation of the beads with glutaraldehyde must be carried out shortly before they are coupled to the antibody.

A mixture was formed consisting of 0.6 mg of spheres suspended in one ml water per 0.1 ml of 50% glutaraldehyde. The mixture was stirred overnight to react the free amino group of the aminododecyl-amido-linked

microspheres with the glutaraldehyde. Excess glutaraldehyde was removed by extensive dialysis.

The anti-H-2K antibody was grown in serum-free media, to prevent contamination with albumin, which has been found to bind to the bead. The antibody is used in the form of supernatant in Dulbecco's (modified) MEM in serum-free media supplemented with 1.5 mg oxalacetate, 0.5 mg insulin, 3.5 mg transferrin, 0.12  $\mu$ g ethanolamine and 0.043  $\mu$ g sodium selenite per liter of DMEM. One ml of unpurified antibody (about 0.1 microgram) was added per 6 mg of microspheres to the glutaraldehyde-activated microspheres. The resulting mixture was stirred for 16 hours at 4°C before glycine was added to react with the remaining unconjugated aldehyde groups. The mixture was layered on a discontinuous sucrose density gradient (60% w/w overlaid with 10-20%) in phosphate buffered saline (PBS), pH 7.4, and spun in a Spinco SW 10.1 rotor ultracentrifuge for one hour at 35,000 rpm. The antibody-microsphere conjugates were recovered at the 20-60% interface and dialyzed overnight against PBS, pH 7.4.

The antibody-microsphere conjugates were found to be primarily in single suspension, with no visible large aggregates or clumps of microspheres.

## EXAMPLE 4

## Use of Antibody/Fluorescent Microspheres To Visualize Cell Surface Antigens

The binding specificity of the antibody-microspheres was determined by incubation with three groups of splenocytes. The first group carried the H-2<sup>d</sup> antigen, the second lacked the relevant H-2<sup>d</sup> antigen, and the third carried the H-2<sup>d</sup> antigen but was preincubated with free monoclonal antibody. The second and third groups served as controls. Following thirty minutes' incubation at room temperature, cells were washed twice with PBS (pH 7.4) and then scored for fluorescence under a Leitz Ortholux fluorescent microscope with an A filter combination consisting of 340-380 m $\mu$  for excitation and 430 m $\mu$  for emission at a magnification of 630 $\times$  to 1,000 $\times$ . A minimum of one hundred cells per group were scored.

When Do4 or Do9 containing fetal bovine serum were used for conjugation under various conditions, the binding efficiency was in the range of 40-60% for group 1 and 14-20% for group 2. By polyacrylamide gel electrophoresis, abundant albumin (more than 98%) was found in supernatant containing bovine serum. Conjugation experiments using albumin alone showed that albumin not only binds to microspheres but also to the cell surface, and thus decreases substantially the antigen-specific binding. An alternative monoclonal antibody (Do4.7) secreted in serum-free media was prepared for the study. Eighty-eight percent, 27% and 4% of binding specificity was detected in group 1, 2 and 3 respectively. However, the 27% binding in the negative control prompted optimizing the method in the following manner:

1. With various concentration of antibody (Ab) to 1 ml of fluorescent microspheres (6 mg/ml) using 1 hour reaction time for conjugation, the maximal binding specificity was obtained using 0.3 ml of Do4.7 antibody per 1 ml of fluorescent microspheres. However, the 20% range of non-specific binding in group 2 remains. This may be caused by inefficient binding of immuno-

globulin, thereby allowing the fluorescent microspheres to bind to the cell surface non-specifically.

2 By studying the time effect on efficiency of the immunoglobulin binding to fluorescent microspheres (0.25 ml of Do4.7 antibody per 1 ml of fluorescent microspheres), the optimal condition to achieve the maximal covalent binding of antibody to fluorescent microspheres was determined to be a four hour reaction. The maximal binding specificity to group 1 is 88% while the non-specific binding for group 2 is reduced to 10-11%.

#### EXAMPLE 5

##### Attachment of Avidin to Fluorescent Microspheres

Avidin was attached to the microspheres of Example 2 by a glutaraldehyde reaction similar to that used in Example 3.

A mixture was formed consisting of 2.4 mg of spheres suspended in four ml water per 0.4 ml of 50% glutaraldehyde. The mixture was incubated for one hour to react the free amino group of the aminododecyl-amido-linked microspheres with the glutaraldehyde. Excess glutaraldehyde was removed by extensive dialysis, overnight against PBS pH 7.4. The glutaraldehyde activated microspheres were concentrated down to 1 ml by using Centricon-30, followed by gently mixing the 1 ml microsphere with 200ul (1 mg/1 ml) streptavidin for four hours. At the end of four hours, 1 mg of lysine was added and further mixed for an additional hour to react with the remaining unconjugated aldehyde groups.

The mixture was layered on a discontinuous sucrose density gradient (60% w/w overlaid with 10-20%) in 1.0M NaCl, 0.01M EDTA pH 8.5 and spun in a Spinco SW 50.1 rotor ultracentrifuge for one hour at 35,000 rpm. The avidin-microsphere conjugates were recovered at the 20-60% interface and dialyzed overnight against 1.0M NaCl, 0.01M EDTA, pH 8.5.

The avidin-microsphere conjugates showed no tendency to clump into visible aggregates. Electron microscopic examination of the beads showed primarily single beads, with occasional groups of two to four beads.

To determine the presence of reactive avidin on the fluorescent microspheres, the avidin fluoromicrosphere complexes were spotted on a nitrocellulose paper in a serial dilution and were detected using a commercially available alkaline phosphatase colorimetric method produced by Bethesda Research Laboratories. It was found that the complex can be detected in a hundred-fold dilution, as compared with a negative control.

#### EXAMPLE 6

##### Use of Avidin/Fluorescent Microspheres To Detect Single Gene Sequences in Plasmid DNA

The specificity of the microspheres produced in accordance with Example 5 for the purpose of detecting gene sequences was determined as follows. Plasmid DNA encoding the N-myc oncogene sequences was first linearized using a restriction enzyme (Eco R1) and denatured. The DNA sequences were then labeled with biotin-16-dUTP using the oligolabeling method of Feinberg and Vogelstein, *Anal. Biochem.*, 132, pp. 6-13 (1983). The biotinylated plasmid DNA was incubated with the avidin/fluoromicrosphere for one hour. The resulting conjugates of DNA-biotin-avidin-microspheres was examined under an electron microscope. It is estimated that more than 175 biotin molecules per thousand base pairs were incorporated. Following denaturation, the single stranded plasmid DNA encoding

N-myc oncogene sequences labeled with biotin-16-dUTP therefore serves as a DNA probe.

#### EXAMPLE 7

##### Use of Avidin/Fluorescent Microspheres To Detect Single Gene Sequences Via Southern Blot

The avidin/fluorescent microspheres of Example 5 may be utilized in a modification of the standard Southern blot procedure, as described for example in Davis, Dibney and Battey, "Basic Methods In Molecular Biology" (Elsevier 1986), pp. 62-65, to detect gene sequences fluorimetrically, rather than radiochemically as in the standard Southern blot procedure.

DNA is extracted from lymphocytes or fibroblast cultures and cleaved with restriction endonucleases at specific sequences. The resulting DNA fragments are separated by gel electrophoresis on the basis of their molecular weight. The size of the fragments can therefore be determined by calibrating the gel with a known size standard. The DNA fragments are then transferred from their position in an agarose gel to a nitrocellulose filter placed directly above the gel. The DNA is denatured, neutralized and transferred in a high-salt buffer by capillary action. The resulting single-stranded DNA binds to the filter and is permanently bonded by baking the filter. This procedure is well known and is described in "Basic Methods In Molecular Biology" supra, pp. 41-61. The bonded DNA is later hybridized to a biotinylated probe as in Example 6, by the method of Feinberg and Vogelstein, supra, to detect hybridizing species. This is followed by incubation with the avidin/fluoromicrospheres of Example 5 onto the nitrocellulose paper for one hour with polylysine. The presence of the probe (the known DNA sequences) can be detected instantly by visualization of the fluorescence when exposed to a portable UV light, or the fluorescence can be quantitated by a spectrofluorometer.

#### EXAMPLE 8

##### Use of Avidin/Fluorescent Microspheres To Detect Gene Sequences Via In Situ Hybridization to the DNA of Chromosomes

As described in the foregoing Examples, single stranded plasmid DNA encoding N-myc or ribosomal gene sequences, or any other gene sequences of interest, labeled with biotin-16-dUTP can be utilized to detect gene sequences via in situ hybridization as follows.

Chromosomes obtained from neuroblastoma cell line, or any other tissues such as lymphocyte and fibroblast, can be first identified with Quinacrine staining (so-called Q-banding) under fluorescence microscopy using the proper filter combinations. In a Leitz fluorescent microscope, H filter combination is used for Quinacrine. Metaphase cells showing Q banding are photographed and their coordinates recorded. The slides are destained and ready to be used for in situ hybridization.

The method used for in situ hybridization may follow that of Harper and Saunders, *Chromasoma*, 83, pp 431-39 (1981), with certain modifications. After thorough washing to remove Q staining, slides of fixed metaphase chromosomes are subjected to the following: removal of endogenous RNA by incubating with ribonucleases; denaturing of chromosomal DNA; hybridization by incubating the previously labeled biotinylated DNA probe on the slide overnight; incubation of the slide with avidin/fluoromicrospheres; examination of fluorescent microspheres attached consistently to the

location of a specific chromosome. The final step is done by first relocating the previously recorded Q-banded chromosome and then detecting the presence of fluoromicrospheres under fluorescent microscopy using A filter (a filter used for examining dansyl fluorescence) and correlating the frequent occurrence of the fluoromicrospheres on a specific location of the previously identified chromosome, thereby mapping the gene to a specific chromosome.

It will be seen from the foregoing examples that the present invention provides fluorescent microspheres which are smaller than previously known fluorescent microspheres, yet which are more intensely fluorescent than even much larger microspheres. They thus permit visualization of far more precisely defined areas of a substrate than previously possible, permit far brighter marking of closely related sites, permit direct localization of cell surface antigens without a secondary antibody, and permit conjugation of DNA to fluorescent microspheres by taking advantage of the strong non-covalent interaction between avidin and biotin. The microspheres may be made with various fluorochromes which include primary amine groups. They are chemically stable for long periods in single suspension. They are thus versatile tools with wide applications for the detection of minute quantities of biological substances via rapid, simple, safe and highly sensitive methodology.

Numerous variations, within the scope of the appended claims, will be apparent to those skilled in the art in light of the foregoing description. For example, automated techniques such as flow cytometry, may be used with the beads. Other fluorescent dyes may be utilized such as rhodamine B, and visible light dyes such as brilliant cresyl blue may also be used. These dyes are preferably, but not necessarily, attached to spacers. They are added in excess (70-90% dye) with the diamine spacers. Biotin may be added to the diamine reactive groups via other means such as N-hydrosuccinimido biotin or by photochemical means, for reaction with avidinated probes, or in a sandwich reaction. These variations are merely illustrative.

I claim:

1. A method of forming a dyed microsphere comprising a first step of forming an acrylic latex bead having on its surface at least five thousand ester terminations, and thereafter a second step of simultaneously reacting said ester terminations on said acrylic latex bead with an amine-terminated dye and with a diamine to form by transacylation a conjugate of said amine-terminated dye and said diamine with said bead.

2. The method of claim 1 wherein said reaction is carried out at a pH of from 10.5 to 13.5.

3. The method of claim 1 including a further step of conjugating a protein to said conjugated diamine.

4. The method of claim 3 wherein said protein is chosen from the group consisting of avidin, immunoglobulins, and receptors.

5. The method of claim 1 wherein said amine-terminated dye and said diamine are present in said second step in a ratio of from three to five parts of amine-terminated dye to one part diamine.

6. The method of claim 1 wherein said amine-terminated dye is a fluorescent amine.

7. The method of claim 6 including a further step of separating said microsphere from unconjugated fluorescent amine and unconjugated diamine.

8. The method of claim 6 wherein said latex bead has a diameter of 200 to 400 angstroms, and wherein said second step of reacting said ester terminations on said latex bead with said fluorescent amine conjugates at least 10,000 fluorescent molecules to the surface of the bead.

9. The method of claim 6 wherein said diamine is a spacer comprising a carbon chain having a length of from eight to twenty carbons, and including a further step of attaching to said spacers a selectively reactive means for selectively reacting with a chemical species on a substrate to label said substrate.

10. The method of claim 8 wherein said fluorescent molecules are conjugated to said beads by carbon chains having a length of from two to ten carbons and wherein said diamines comprise spacers including carbon chains having a length of from eight to twenty carbons, said second step of simultaneously reacting said ester terminations on said latex bead with said fluorescent amine and diamine comprising reacting said fluorescent amine and said diamine in a ratio of from three to five parts of fluorescent amine to one part diamine.

11. The method of claim 1 including a further step of separating said microsphere from unconjugated amine-terminated dye and unconjugated diamine.

12. The method of claim 11 wherein said latex bead has a diameter of 200 to 400 angstroms, and wherein said second step of reacting said ester terminations on said latex bead with said amine-terminated dye bead conjugates at least 10,000 dye molecules to the surface of the bead.

13. The method of claim 12 wherein said diamine is a spacer comprising a carbon chain having a length of from eight to twenty carbons, and including a further step of attaching to said spacers a selectively reactive means for selectively reacting with a chemical species on a substrate to label said substrate.

14. The method of claim 13 wherein said dye molecules are conjugated to said beads by carbon chains having a length of from two to ten carbons and wherein said diamines comprise spacers including carbon chains having a length of from eight to twenty carbons, said second step of simultaneously reacting said ester terminations on said latex bead with said amine-terminated dye and diamine comprising reacting with amine-terminated dye and said diamine in a ratio of from three to five parts of amine-terminated dye to one part diamine.

15. The method of claim 1 wherein said ester terminations include both alkoxy groups and alkoxy alcohol groups which react with the amine-terminated dye and the diamine.

16. The method of claim 1 including a further step of attaching to said diamine a protein or protein conjugate by reacting the diamine with a dialdehyde to produce an aldehyde termination and thereafter reacting the aldehyde termination with said protein or protein conjugate.

17. The method of forming a fluorescent microsphere comprising forming a latex bead, thereafter a step of chemically attaching to said latex bead a first set of spacers having attached thereto fluorescent labels and simultaneously chemically attaching to said latex bead a second set of spacers, and thereafter a step of chemically attaching to said second set of spacers a selectively reactive means for selectively reacting with a chemical species on a substrate to label said substrate.

18. The method of claim 17 wherein said second set of spacers include amine terminations, and said step of

**11**

attaching to said second set of spacers a selectively reactive means comprises attaching said selectively reactive means by reacting said amine terminations with a dialdehyde to produce aldehyde terminations, and

**12**

thereafter reacting the aldehyde terminations with said selectively reactive means.

19. The method of claim 17 wherein said selectively reactive means is an avidin.

20. The method of claim 17 wherein said selectively reactive means is an antibody of antigen.

\* \* \* \* \*

10

15

20

25

30

35

40

45

50

55

60

65